Exhibit B

ASSAY FOR THE SECRETED ALKALINE PHOSPHATASE (SEAP)

1> - From the 48-hour cotransfected cell cultures, remove 250 µl of each culture supernatant and transfer into Eppendorf tubes. To be on the safe side, maintain the cultures at 37°C until satisfactory data have been obtained.

2> - Heat samples at 65°C for 5 minutes to inactivate endogenous phosphatases (Seap is stable at 65°C). homoarginine also inactivate endogenous phosphates

3> - Centrifuge for 2 minutes at room temperature in a Microfuge.

4> - Transfer 150 µl supernatants to new Eppendorf tubes. At this stage, samples may be stored at -20°C indefinately.

5> - In an Eppendorf tube, add 100 µl of 2 ~~x Seap buffer~~ to 100 µl of sample. As a Zero standard make up a mixture in triplicate substituting sample with water. Mix on a Vortex.

6> - Transfer the contents of each tube to a well of a flat bottom microtiterplate. Avoid creating air bubbles.

7> - Incubate plate at 37°C for 10 - 15 minutes.

8> - During this incubation make up the *p*-nitrophenylphosphate solution (Seap enzyme substrate) and prewarm it at 37°C for 5 minutes.

9>- Add 20 µl of the substrate solution to each well, preferably using a multipipetter.

10>- Using an ELISA microplate reader with an automatic shaker and incubator unit, measure the OD at 405 nm at regular intervals (e.g. every 5 minutes) over 60 minutes while the plate is being incubated at 37°C. (Program a 5-seconde shaking before any reading).

11>- Calculate the levels of Seap activity at a point on the curve when the changes in OD are linear with respect to time (e.g. at 15 - 30 minutes).

********** * **********

Buffer and Chemicals

2 x Seap Buffer (for 50 ml)

Amount	Stock	Final Conc.
10.51 g*	diethanolamine (100% sol.)	2 M
50 µl	1 M $MgCl_2$	1 mM
226 mg	L-homoarginine	20 mM

(*) Weigh exactly 10.51 g of diethanolamine in a tared beacker. Add distilled water up to 45 ml. Stir to homogenize. Add 50 µl of 1 M MgCl2 while stirring. In a separate 15-ml conical tube, dissolve 226 mg of L-homoarginine in 2-3 ml of distilled water. Add the solution to diethanolamine/MgCl2 solution under constant stirring. Bring up to 50 ml.

120 mM p-nitrophenylphosphate is made in 1 x Seap buffer (fresh).

Amount G (mg) = (120 mM x 371.12 x vol)/1000

Where: vol = [(# wells x 20 µl) + 100 µl extra]/1000

* Make the solution in 1 x Seap buffer (make fresh)

For 51 wells of the microplate (48 samples + 3 blanks), dissolve 50 mg *p*-nitrophenylphosphate in 1.120 ml of 1 x Seap buffer. → 600 ml stock + 600 µl H_2O

Chemicals

Name	Cat No	Storage
Diethanolamine (Fluka)	31589	Room T
L-homoarginine hydrochloride (Sigma)	H-1007	4°C
p-nitrophenylphosphate* (Fluka)	71768	4°C

(*) Also known as 4-nitrophenylphosphate disodium salt hexahydrate

0.218 g
0.226 "

SEAP ASSAY SHEET

I - Assay Title:

* Assay #: 72 Date #: [redacted] Investigator's Name: Wen

*Test Compounds: Mal 4 & Mal.4 3Na+ transfected at [redacted] 11:30

*Concentrations:

II - DNA Transfection: Ratio (2:1) HIV/SEAP:pcTAT

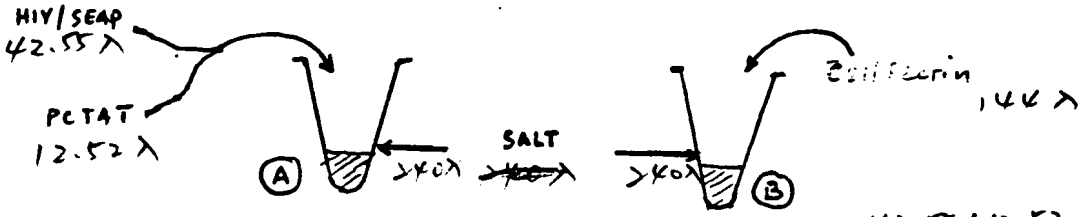
*HIV/Seap: 0.4 µg/well x40 = 16.0 mg ===> From 0.376 % stock: 42.55 λ

*pcTAT: 0.2 µg/well x40 = 8.0 mg ===> From 0.639 % stock: 12.52 λ

*Total DNA (µg) = 24 mg

* Vol. Cell Fectin = Total DNA x 6 = 144 λ

* Vol. 150 mM NaCl = Cellfectin (µl) / 0.6 = 240

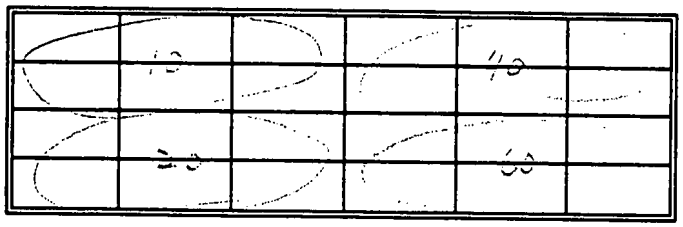


* Transfection cocktail (µl)/ well: (240+240+144+42.55+12.52)/40 = 16.98 λ

III. Linbro® 24 flat bottom well of 17 mm

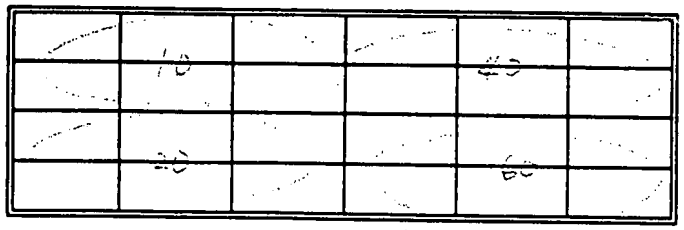
A

Mal 4 DMSO



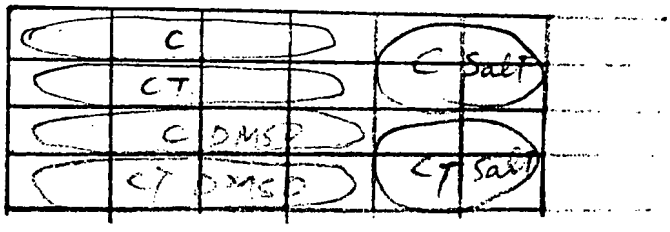
B

Mal 4 Salt



Preparation of Drug conc.

C (NOTE)



40 : 98
60 : 97

40 : 23
60 : 46

REPEATED-READS #: 0025

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TABLE OF ABSORBANCE VALUES

FILE: FILE 3 TITLE: FILE 3 REPORT

	1	2	3	4	5	6	7	8	9	10	11	12
A	0#000	-0#002	0#004	-0.217	-0.215	-0.216	0.064	0.080	0.089	0.126	0.133	0.146
B	-0.210	-0.214	-0.205	-0.215	-0.215	-0.213	0.151	0.153	0.207	0.124	0.153	0.155
C	0.070	0.016	0.060	0.044	-0.214	-0.214	0.105	0.089	0.131	0.362	0.171	0.169
D	0.689	0.649	0.663	0.756	-0.208	-0.210	0.126	0.122	0.120	0.352	0.172	0.197
E	0.094	0.100	0.081	0.110	-0.212	-0.209	0.093	0.067	0.087	0.101	0.080	0.100
F	0.680	0.711	0.680	0.614	-0.211	-0.213	0.830	0.776	0.832	0.649	0.587	0.501
G	0.064	0.067	0.080	0.079	-0.203	-0.211	0.089	0.114	0.100	0.114	0.118	0.098
H	0.784	0.708	0.643	0.684	-0.209	-0.205	0.681	0.720	0.609	0.481	0.437	0.437

Mal.4 DMSO

C 0.048
CT 0.689
0.641
C DMSO 0.096
CT DMSO 0.671
0.575
C NaOH 0.073
CT NaOH 0.705
0.632
Mal.4 0 ug/mL 0.078
Mal.4 10 ug/mL 0.170
Mal.4 20 ug/mL 0.108
Mal.4 40 ug/mL 0.135
Mal.4 60 ug/mL 0.144
Mal.4 20 ug/mL 0.123
Mal.4 60 ug/mL 0.185
Mal.4 10 ug/mL 0.082
Mal.4 40 ug/mL 0.094
Mal.4 10 ug/mL 0.813
Mal.4 40 ug/mL 0.579
Mal.4 10 ug/mL 0.101
Mal.4 20 ug/mL 0.110
Mal.4 60 ug/mL 0.452
Mal.4 20 ug/mL 0.690

Mal.4 - DMSO Ref: DMSO
% inhibition
10 ug/mL : 84
20 ug/mL : 97
40 ug/mL : 98
60 ug/mL : 97

Mal.4 - NaOH Ref: NaOH
% inhibition
10 ug/mL : 0 (-16)
20 ug/mL : 10
40 ug/mL : 23
60 ug/mL : 46

SEAP assay of NDGA derivatives [redacted]

1. Detach COS7 cells from four 90mm culture plates by adding 0.05% Trypsin (0.05% Trypsin in CMF-PBS, 1mM EGTA) 0.5 ml after 2 times CMF-PBS wash.

2. Inactivate trypsin by adding 5ml complete IMDM medium (10% Fetal Bovine Serum, antibiotics) to each plate.

3. Suspend the cells by gentle pipetting. Count by Hemocytometer 160×10^4 cells/cm^3 is the cell density.

4. Seed 80ul cell suspension into 10 Linbro 24-wells culture plates which were precoated with 0.1% gelatin & contained 0.3ml complete medium each well. (1.3×10^5 cells/well) ([redacted] 18:00)

5. Transfect the cells after 26h incubation (37°C. 95% Air- 5% CO_2) by Adding 30 ul ppt (0.6 ug DNA/well) to each well that containing 0.3 ml fresh complete medium. The control wells accept same ppt without DNA. ([redacted] 20:00)

Ca-PO4 ppt preparation:

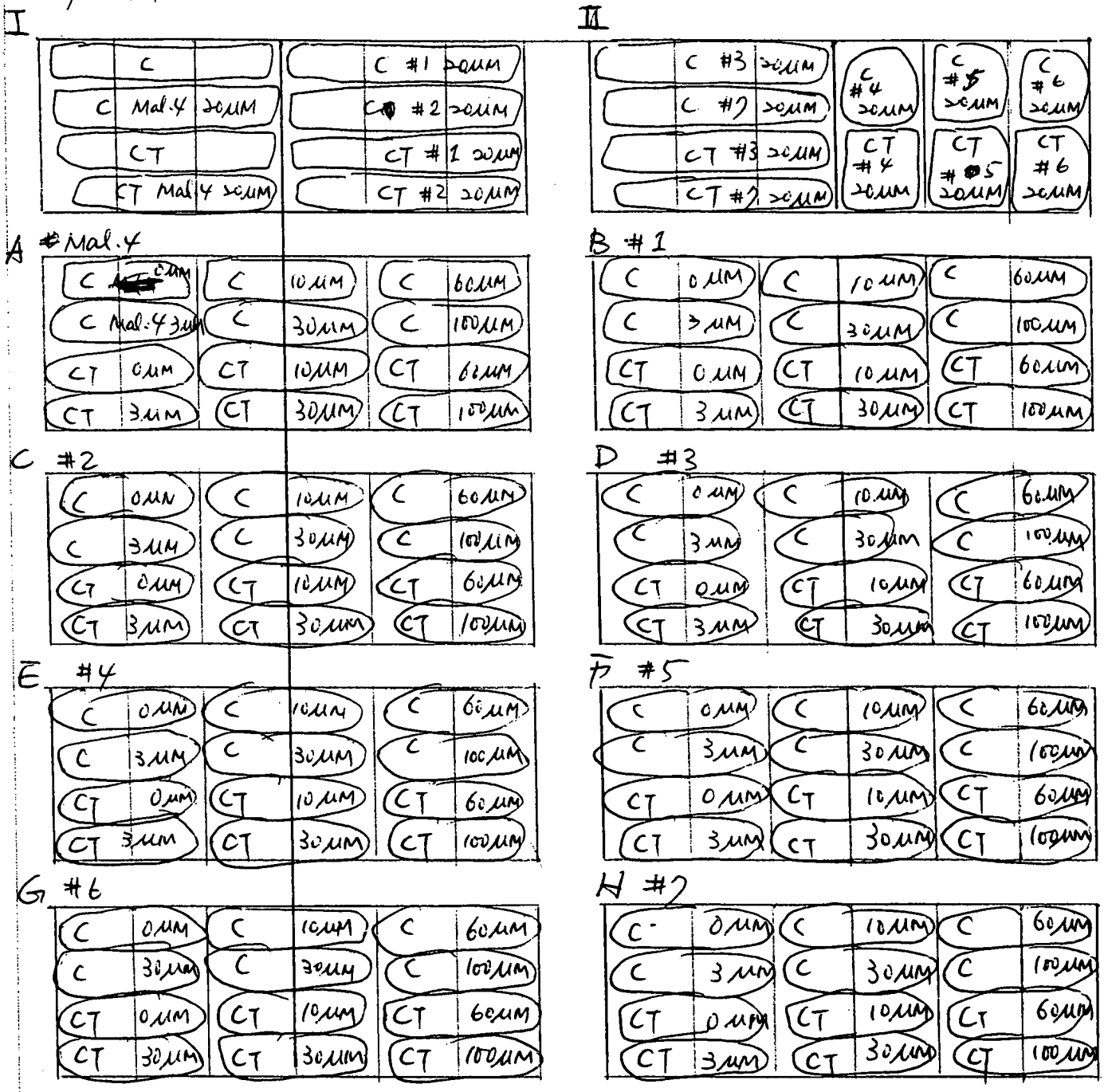
a. for control wells: Ca^{2+} soln (0.25M $CaCl_2$, 0.025M HEPES, pH 7.1) 1ml drop into 1ml bubbling PO_4^{2-} soln (0.28M NaCl, 0.025M HEPES, 0.0015M Na_2HPO_4, pH 7.1).

b. for transfect wells:

Three 2-ml tubes containing ~~pBC12/HIV/SEAP 18ug, pBC12/CMV/t2 9ug,~~ 675ul PO_4^{2-} soln were added Ca^{2+} soln 675 ul ~~on~~ with DNAs (pBC12/HIV/SEAP 18ug, pBC12/CMV/t2 9 ug) dropwise with bubbling. then Sit 30 min before use.

6. Incubate at 37°C for 18h.

7
6 Map of Plates:



C = Control wells (without DNA)
CT = Transfect wells (with DNA)

8 Remove growth medium, add 0.500ul medium containing test compounds. The final concentration of DMSO is 0.3% ([redacted]).
△ Dilution of test compounds:

Stock: MonoMe: 10.53 ug/ul use 3ul in 1ml medium = 100uM
DiMe: 11.0 ug/ul "
TriMe: 11.47 ug/ul "
tetraMe: 11.93 ug/ul "

For lower concentration, dilute the stock by DMSO, & use 3ul in 1ml medium to keep the final DMSO concentration is 0.3%.
△ DMSO stock was ~~add~~ mixed into medium just piror to use by vortexing.

9 Incubate for 48h. Remove 200ul medium from each well. for SEAP assay. ([redacted] 14:00).

10 The medium used in SEAP assay is 10ul for each sample.

MEASMTEK-READER AT 0625

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TABLE OF ABSORBANCE VALUES

FILE: FILE 3 TITLE: FILE 3 REPORT

	1	2	3	4	5	6	7	8	9	10	11	12
A	-0#024	0#005	0#020	-0.217	-0.216	-0.217	0.011	0.012	0.013	0.004	-0.005	-0.002
B	0.012	-0.008	0.012	0.012	0.020	0.006	0.002	0.004	0.005	0.003	0.019	0.003
C	-0.004	0.004	0.016	0.004	0.008	-0.001	0.585	0.704	0.631	0.558	0.394	0.421
D	1.126	1.040	1.102	0.801	0.766	0.811	0.347	0.373	0.326	0.502	0.428	0.410
E	1.010	0.855	0.915	0.890	0.753	0.799	0.018	0.020	0.007	0.012	-0.006	0.023
F	-0.213	-0.216	-0.215	-0.213	-0.213	-0.216	0.008	0.014	0.007	-0.010	0.010	0.036
G	0.007	0.011	0.013	0.005	0.001	0.017	1.323	1.433	1.105	1.111	0.292	0.269
H	0.007	-0.001	0.016	0.003	0.009	0.026	1.284	1.234	0.783	0.715	0.165	0.162

I II A B C D

Mal.% % inhibition

3 mM = 8.2

10 mM = 19.2

30 mM = 44.7

60 mM = 80.0

100 mM = 89.6

30 mM inhibition

Mal.% : 14.9

1 : 28.0 =

2 : 25.3 =

3 : 42.1 =

4 : 51.5 =

5 : 62.8 =

6 : 61.7 =

7 : 68.2 =

REPLICATED-READS # • 0025

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TABLE OF ABSORBANCE VALUES

FILE: FILE 3 TITLE: FILE 3 REPORT

	1	2	3	4	5	6	7	8	9	10	11	12
A	0#002-0#004		0#003-0.211	-0.209	-0.212		0.008	0.016	0.003	-0.001	0.002	0.016
B	0.002	0.002	0.012	0.012	0.015	0.006	0.003	0.009	0.000	-0.003	0.010	0.009
C	-0.005	0.007	0.008	0.004	0.007	-0.001	1.200	1.149	0.871	1.088	0.374	0.357
D	1.385	1.306	1.025	1.098	0.423	0.464	1.067	1.185	0.570	0.530	0.236	0.226
E	1.288	1.327	0.905	1.005	0.330	0.314	0.017	0.012	0.002	0.007	0.003	0.016
F	-0.202	-0.210	-0.210	-0.210	-0.209	-0.209	0.006	0.010	0.009	0.002	0.004	0.010
G	1.239	1.354	1.209	1.239	0.420	0.350	1.122	1.205	0.921	0.933	0.208	0.218
H	1.387	1.279	0.682	0.580	0.203	0.193	1.178	1.165	0.475	0.435	0.085	0.114

#1 % inhibition

3 uM : -3.3(0)
10 " : 5.7
30 " : 52.8
60 " : 70.8
100 " : 86.0

#2 % inhibition

3 uM : 2.8
10 " : >1.9
30 " : >9.4
60 " : 67.8
100 " : 76.3

#3 % inhibition

3 uM : 3.7
10 uM : 15.8
30 uM : 52.5
60 uM : 69.3
100 uM : ~~80~~ 81.0

#4 % inhibition

3 uM : -1.3(0)
10 uM : 19.8
30 uM : 60.9
60 uM : 82.4
100 uM : 92.2

KLPEATED-READS #: 0025

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TABLE OF ABSORBANCE VALUES

FILE: FILE 3 TITLE: FILE 3 REPORT

	1	2	3	4	5	6	7	8	9	10	11	12
A	0#000	-0#008	0#010	-0.210	-0.208	-0.210	-0.209	-0.209	-0.210	-0.208	-0.209	-0.209
B	-0.207	-0.208	-0.209	-0.208	-0.208	-0.209	-0.208	-0.209	-0.209	-0.208	-0.209	-0.211
C	0.012	0.014	0.012	0.022	0.017	0.020	0.011	0.014	0.011	0.014	0.019	0.015
	0.013		0.017		0.019		0.013		0.013		0.017	
D	0.007	0.013	0.015	0.014	0.026	0.012	0.010	0.019	0.004	0.020	0.038	0.043
	0.010		0.015		0.019		0.015		0.012		0.041	
E	1.192	1.134	0.699	0.687	0.277	0.262	1.391	1.489	0.658	0.623	0.317	0.300
	1.163		0.693		0.270		1.440		0.641		0.309	
F	1.215	1.138	0.374	0.400	0.202	0.184	1.163	0.974	0.380	0.317	0.189	0.239
	1.177		0.387		0.193		1.069		0.349		0.214	
G	-0.205	-0.208	-0.208	-0.206	-0.199	-0.208	-0.206	-0.207	-0.207	-0.207	-0.209	-0.210
H	-0.207	-0.207	-0.204	-0.206	-0.206	-0.204	-0.189	-0.207	-0.207	-0.206	-0.206	-0.210

G #6

H #7

#6
% inhibition

3 μM = -1.5 (0)
10 μM = 41.0
30 μM = 67.7
60 μM = 78.2
100 μM = 84.9

#7
% inhibition

3 μM : 26.1
10 μM = 56.0
30 μM = 76.4
60 μM = 79.5
100 μM = 87.9

REPEATED-READS #: 0025

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TABLE OF ABSORBANCE VALUES

FILE: FILE 3 TITLE: FILE 3 REPORT

	1	2	3	4	5	6	7	8	9	10	11	12
A	-0#005	-0#012	0#018	-0.209	-0.210	-0.211	-0.210	-0.209	-0.208	-0.209	-0.210	-0.211
B	-0.208	-0.209	-0.209	-0.209	-0.209	-0.209	-0.209	-0.209	-0.209	-0.209	-0.207	-0.211
C	0.028	0.031	0.037	0.037	0.040	0.041	-0.206	-0.207	-0.209	-0.208	-0.208	-0.209
D	0.015	0.028	0.036	0.028	0.047	0.035	-0.208	-0.210	-0.208	-0.208	-0.208	-0.209
E	1.500	1.394	0.701	0.680	0.318	0.291	-0.207	-0.207	-0.208	-0.207	-0.209	-0.209
F	1.131	1.033	0.385	0.417	0.222	0.185	-0.207	-0.208	-0.207	-0.206	-0.208	-0.213
G	-0.206	-0.206	-0.208	-0.207	-0.206	-0.208	-0.206	-0.205	-0.207	-0.207	-0.208	-0.208
H	-0.207	-0.204	-0.209	-0.208	-0.207	-0.207	-0.207	-0.207	-0.207	-0.207	-0.207	-0.207

#5 B

C: 0.030, 0.039, 0.041

D: 0.022, 0.032, 0.041

E: 1.447, 0.691, 0.305

F: 1.082, 0.401, 0.204

#5 % inhibition

3 uM : 25.2

10 uM : ~~53.8~~ 53.8

30 uM : 74.0

60 uM : 81.4

100 uM : 88.5

% Inhibition of NDGA Derivatives on SEAP assay.

	Mal.4	#1	#2	#3	#4	#5	#6	#7
3 μM	8.2	0 (-3.3)	2.8	3.7	0 -1.3	25.2	0-1.5	26.1
10 μM	19.2	5.7	21.9	15.8	19.8	53.8	40.1	56.0
30 μM	44.7	~~8.8~~ 52.8	29.4	52.5	60.9	74.0	67.7	76.4
60 μM	80.0	~~53.8~~ 70.8	67.8	69.3	82.4	81.4	78.2	79.5
100 μM	89.6	86.0	76.3	81.0	92.2	88.5	84.9	87.9

Figure B(1)

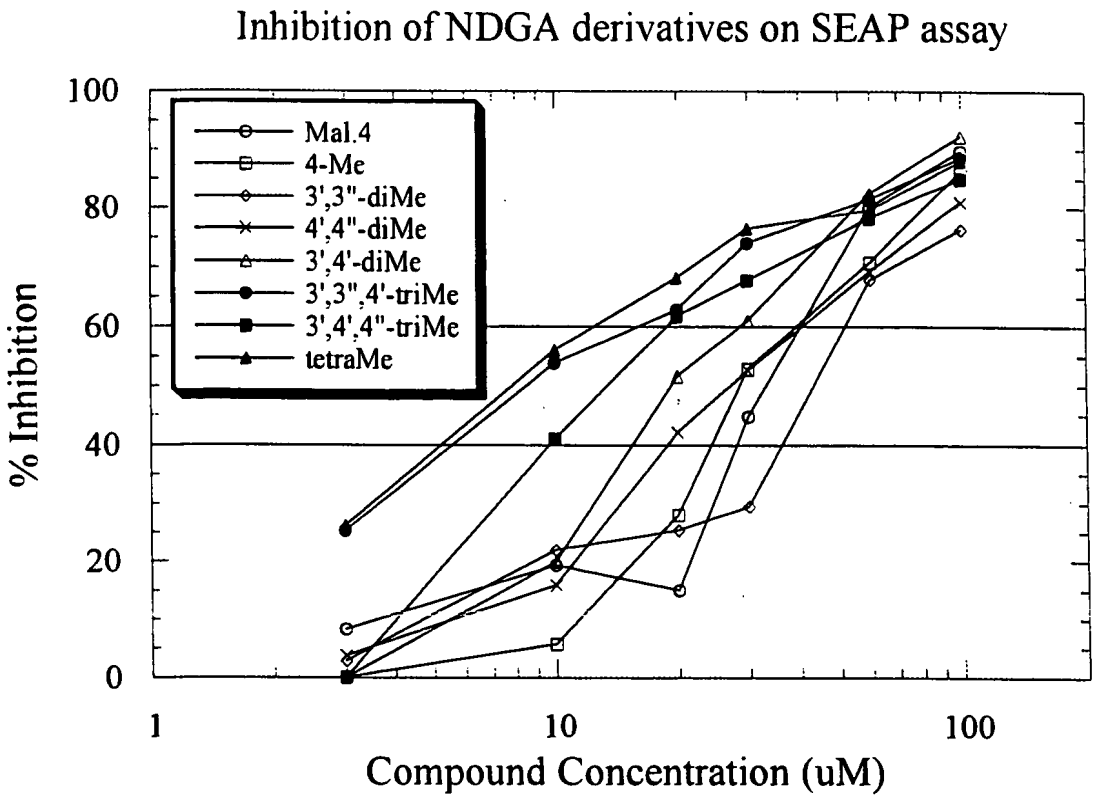

Figure B(2)